

Full-length article

Pharmacokinetics of His-tag recombinant human endostatin in Rhesus monkeys¹

Hai-feng SONG², Xiu-wen LIU, Hai-ning ZHANG, Bao-zhen ZHU, Shou-jun YUAN, Shang-yi LIU, Zhong-ming TANG*Department of Pharmacology and Toxicology, Beijing Institute of Radiation Medicine, Beijing 100850, China*

Key words

endostatin; pharmacokinetics; *Macaca mulatta*; immunoenzyme techniques¹ Project supported by the National Natural Science Foundation of China, No 39930180.² Correspondence to Assoc Prof Hai-feng SONG.

Phn 86-10-6693-1230. Fax 86-10-6693-1230. Email songhf@nic.bmi.ac.cn

Received 2004-05-12

Accepted 2004-08-03

doi: 10.1111/j.1745-7254.2005.00009.x

Abstract

Aim: To study the pharmacokinetics and accumulation of an *Escherichia coli*-expressed His-tag fused recombinant human endostatin (rh-endostatin) in Rhesus monkeys. **Methods:** Rh-endostatin was iv or sc injected in Rhesus monkeys, and the rh-endostatin concentration in serum samples was determined by an enzyme immunoassay (EIA) method. The serum drug concentration-time data were analyzed by compartmental analysis using the practical pharmacokinetic program 3p97. **Results:** Following iv administration at a dose rate of 1.5, 4.5, and 13.5 mg/kg in rhesus monkeys, the concentration-time curves of rh-endostatin were best fitted to a three-compartment open model. $AUC_{(0-\infty)}$ linearly increased with dose, while Cl_s exhibited no significant difference among different dose groups. The terminal half-lives (λ_3) were 8 ± 8 , 3.1 ± 1.4 , and 20 ± 14 h, respectively. After sc administration at a dose rate of 1.5 mg/kg, the concentration-time curve was best fitted to a two-compartment open model, with a terminal half-life ($T_{1/2\beta}$) of 8 ± 3 h. Bioavailability following sc injection was approximately $70\%\pm 3\%$. After consecutive iv injection of rh-endostatin at a dose rate of $1.5\text{ mg}\cdot\text{kg}^{-1}\cdot\text{d}^{-1}$ for 7 d, the $AUC_{(0-24\text{ h})}$ substantially increased from $22\pm 13\text{ mg}\cdot\text{h}\cdot\text{L}^{-1}$ (d 1) to $50\pm 29\text{ mg}\cdot\text{h}\cdot\text{L}^{-1}$ (d 7), with an accumulation factor of 2.3 ± 0.6 ($P<0.05$). **Conclusion:** The pharmacokinetic behavior of rh-endostatin in Rhesus monkeys complies with linear kinetics within the examined dose range. It tends to be accumulated in bodies after repeated administration at a dose level of $1.5\text{ mg}\cdot\text{kg}^{-1}\cdot\text{d}^{-1}$ for more than 7 consecutive days.

Introduction

Endostatin, which contains 178 amino acid residues, is a 20 kDa COOH-terminal fragment of collagen XVIII^[1]. It was firstly identified as an angiogenesis inhibitor in 1997. Endostatin specifically inhibits endothelial proliferation and potently inhibits angiogenesis and tumor growth^[2]. Subsequently, *Escherichia coli*-derived recombinant endostatin was shown to induce regression and prevent growth of experimental tumors in mice when administered in daily doses as high as $20\text{ mg}/\text{kg}$ ^[2,3]. A novel *Escherichia coli* derived rh-endostatin was developed by MedGen Ltd (Yantai City, China). Compared with the rh-endostatin reported in the literature, an additional nine-amino acid-sequence (MGGSHHHHH) was added at the N-terminal of

the protein, which resulted in the formation of a six-histidine tag (His-tag). The His-tag can be chelated with metal ions such as Ni^{2+} with a relatively high affinity, so it was utilized to simplify purification and to improve stability of the protein. The present study attempted to reveal the pharmacokinetics and the possibility of accumulation after single or multiple doses of this novel rh-endostatin in Rhesus monkeys.

Materials and methods

Drugs and reagents The rh-endostatin used in this study was provided by MedGen (lot YH0502, purity >98%). The drug was dissolved in 5 mmol/L Tris·HCl (pH 7.2) and stored at 4 °C before use.

The ACCUCYTE[®] Human Endostatin[™] enzyme immunoassay (EIA) kit was purchased from CytImmune Sciences Inc (lot 5AE002-SA, Maryland, USA). All other reagents were purchased from commercial sources and were of analytical grade.

Animals Rhesus monkeys (weighing 5.3 kg±0.8 kg, Grade I, Certificate BDW95002) were provided by the Animal Raising Center of the Academy of Military Medical Sciences, China. The animals were individually housed in stainless-steel cages and fed with a standard monkey diet. Fresh fruit was provided twice per day and water was available *ad libitum*.

Experimental design Three iv groups (at a dose rate of 1.5, 4.5, and 13.5 mg/kg body weight, respectively) were assigned to a 3×3 crossover design to perform an auto-control experiment, in which each animal received the three doses mentioned above, with an interval of 7 d between doses (washing out period). Three animals in the sc group were administered rh-endostatin at the dose rate of 1.5 mg/kg. Finally, a multiple-dose group comprising three animals was also designed, in which the monkeys received seven consecutive iv injections of rh-endostatin at the dose rate of 1.5 mg/kg, once per day.

Sample collection and assay Whole blood samples were drawn from the femoral veins of the animals using a puncture needle immediately before administration and at 2, 10 min in the iv group and 20, 40 min, 1, 1.5 h in the sc group, then at 2, 4, 8, 12, 24, 36, 48, and 72 h in all groups after a single dose and the 1st and the 7th dose in the multiple-dose group. In the multiple-dose group, samples were also collected at 2 min and 24 h after the 4th dose. The blood was kept undisturbed at room temperature, then centrifuged at 3000×g for 10 min. The serum was separated and stored at -20 °C until the assay.

The EIA kit was utilized to determine the rh-endostatin levels in the serum samples. The assay was performed according to the guidance of the procedure specified by the manufacturer. A series of calibration standards provided by Medgen was set up in each microplate. The rh-endostatin levels in the unknown samples were obtained by being calculated from the Medgen calibration standard curves on the same microplate.

Data analysis The concentration-absorbance curves were drawn with the MicroCal Origin software (ver 5.0), and then the experimental data were fitted by Logistic function according to the following formula:

$$Y = \frac{(E_{\max} - E_{\min})}{(1 + (X/EC_{50})^{\text{Slope}})} + E_{\min}$$

In which, “X” represents rh-endostatin concentration in samples, “Y” represents the absorbance at 492 nm. The E_{\max} and E_{\min} were the maximal and the minimal absorbance, respectively.

The pharmacokinetic parameters were obtained by the computer program 3p97. Statistical inference was obtained by means of Student’s *t* test or Chi-square test.

Results

Validity of the method for determination of rh-endostatin in monkey serum The ACCUCYTE[®] Human Endostatin[™] kit has been developed using rh-endostatin as the antigen for polyclonal antibody production, and is used in the kit as the assay standard. Studies have demonstrated that it has no cross-reactivity against a variety of cytokines and growth factors, including heat inactivated rh-endostatin and mouse endostatin.

Within the concentration range of 7.8–8000 µg/L, the concentration-absorbance behavior of the rh-endostatin standard provided by Medgen is presented as an inhibitory sigmoid curve that could be described by 4 parameter logistic fitting. Maximal coefficients of variation (CV%) of intra-assay and inter-assay were 13.7% and 20.8%, respectively. The limit of quantitation (LOQ) was 125 µg/L (with CV% of 6.1%). The average endogenous endostatin level observed in the rhesus monkey was 375±175 µg/L ($n=12$).

The results of the blank serum samples fortified with 2000, 500, and 125 µg/L tested rh-endostatin showed that the recovery rates were 87%±5% (81%–91%), 93%±18% (79%–114%), and 94%±4% (90%–99%), respectively ($n=5$ parallel experiments). The CV% of the intra-assay was less than 11.5%, and the CV% of the inter-assay was higher (as high as 19.7%), so it is important to calculate the rh-endostatin concentration in samples by the parameters obtained from the standard curve of the same microplate.

The validity studies demonstrated that the ACCUCYTE[®] Human Endostatin[™] kit was reliable for the determination of serum endostatin levels. The specificity, sensitivity, accuracy, and precision all met the requirements for pharmacokinetics (PK) study.

Concentration-time curves and PK following single dose of rh-endostatin Serum concentration-time profiles of rh-endostatin following iv bolus injections at doses of 1.5, 4.5, and 13.5 mg/kg were best fitted to a three-compartment open model (Figure 1). Rh-endostatin concentration in serum dramatically decreased after iv injection, with the initial half lives (λ_1) of 0.027±0.016 h, 0.04±0.03 h, and 0.10±0.04 h ($P<0.05$ vs the other two groups) following iv

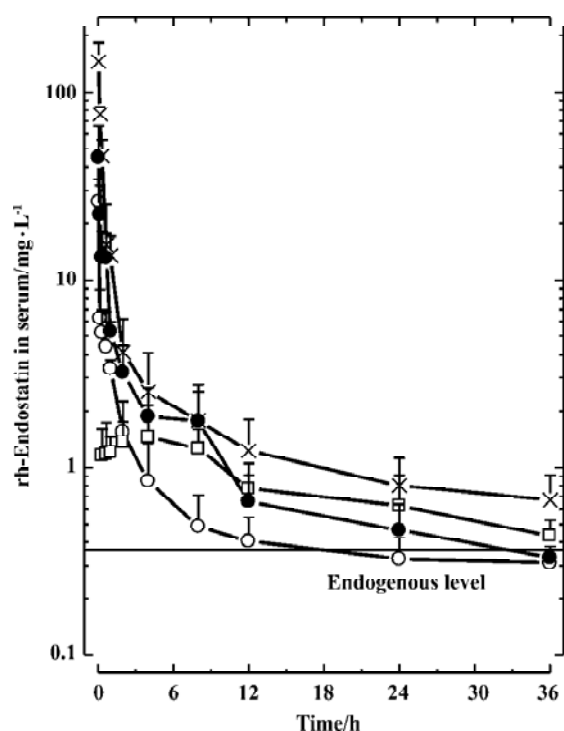


Figure 1. Concentration-time curves of rh-endostatin after a single iv and sc injection in rhesus monkeys. ○, iv 1.5 mg/kg; ●, iv 4.5 mg/kg; ×, iv 13.5 mg/kg; □, sc 1.5 mg/kg. The endogenous endostatin level set at $375 \pm 175 \mu\text{g/L}$.

injection of 1.5, 4.5, and 13.5 mg/kg, respectively. The half-lives of the terminal elimination phase (λ_3) were 8 ± 8 h, 3.1 ± 1.4 h, and 20 ± 14 h, respectively. At a high dose rate of 13.5 mg/kg,

the value of λ_3 was obviously prolonged but without any statistically significant differences. Increased with dose, the mean areas under the serum concentration-time curve [$\text{AUC}_{(0-\infty)}$] of the three iv groups were $15 \pm 5 \text{ mg}\cdot\text{h}\cdot\text{L}^{-1}$, $33 \pm 7 \text{ mg}\cdot\text{h}\cdot\text{L}^{-1}$ and $93 \pm 25 \text{ mg}\cdot\text{h}\cdot\text{L}^{-1}$, respectively. With a dose ratio of 1:3:9, the ratio of $\text{AUC}_{(0-\infty)}$ in the groups was 1:2.2:6.1.

Despite the increase in $\text{AUC}_{(0-\infty)}$, the systemic clearances (Cl_s) among the iv groups were not significantly different (Table 1), indicating that the pharmacokinetic behavior of endostatin within the examined dose range is of linear profile.

The concentration-time profiles following sc administration at a dose rate of 1.5 mg/kg can be described approximately by a two-compartment model (Figure 1). After injection, rh-endostatin was absorbed rapidly, with the time to peak (T_{max}) of 1.1 ± 0.8 h. The mean peak serum concentration (C_{max}) and $\text{AUC}_{(0-\infty)}$ were $1.3 \pm 0.4 \text{ mg/L}$ and $11 \pm 4 \text{ mg}\cdot\text{h}\cdot\text{L}^{-1}$, respectively. The terminal half-life ($T_{1/2\beta}$) was 8 ± 3 h. Compared with the iv group at the same dose, rh-endostatin had a relatively high bioavailability (F) of 70% after sc administration (Table 1).

PK after multiple dosing Following repeated iv administration of rh-endostatin for a 7-d period (once daily) at a dose rate of 1.5 mg/kg, most of the rh-endostatin concentrations at the same time points following injection at d 1 and d 7 were with statistically significant difference (Figure 2). The trough level increased from $388 \pm 61 \mu\text{g/L}$ (d 1) to $623 \pm 261 \mu\text{g/L}$ (d 4) and $1008 \pm 892 \mu\text{g/L}$ (d 7), and the $\text{AUC}_{(0-24\text{h})}$ also increased from $22 \pm 13 \text{ mg}\cdot\text{h}\cdot\text{L}^{-1}$ (d 1) to $50 \pm 29 \text{ mg}\cdot\text{h}\cdot\text{L}^{-1}$ (d 7), but the difference was not significant ($P = 0.053$). The calculated accumulation factor ($\text{AUC}_{\text{d}7}/\text{AUC}_{\text{d}1}$)

Table 1. Pharmacokinetic parameters of the rh-endostatin after iv and sc injection in rhesus monkeys. $n=3$. Mean \pm SD. ^b $P < 0.05$, ^f $P < 0.001$ vs the iv 1.5 mg/kg group.

Parameters	iv			sc
	1.5 mg/kg	4.5 mg/kg	13.5 mg/kg	1.5 mg/kg
$V_c/\text{L}\cdot\text{kg}^{-1}$	0.030 ± 0.019	0.08 ± 0.04	0.078 ± 0.019^f	1.3 ± 0.4^f
$T_{1/2\text{ka}}/\text{h}$	-	-	-	0.34 ± 0.14
$T_{1/2\lambda_1}/\text{h}$	0.027 ± 0.016	0.04 ± 0.03	0.10 ± 0.04^b	-
$T_{1/2\lambda_2}/\text{h}$	0.80 ± 0.28	0.5 ± 0.4	0.8 ± 0.5	1.9 ± 2.0
$T_{1/2\lambda_3}/\text{h}$	8 ± 8	3.1 ± 1.4	20 ± 14	8 ± 3
T_{max}/h	-	-	-	1.1 ± 0.8
$C_{\text{max}}/\text{mg}\cdot\text{L}^{-1}$	-	-	-	1.3 ± 0.4
$\text{AUC}_{0-\infty}/\text{mg}\cdot\text{h}\cdot\text{L}^{-1}$	15 ± 5	33 ± 7^b	93 ± 25^b	11 ± 4
$\text{MRT}_{0-\infty}/\text{h}$	3.6 ± 0.9	2.7 ± 1.5	9.2 ± 2.3^b	9.3 ± 1.7
$\text{CL}_s/\text{L}\cdot\text{h}^{-1}\cdot\text{kg}^{-1}$	0.11 ± 0.03	0.046 ± 0.010	0.017 ± 0.004	0.15 ± 0.05
$V_{\text{ss}}/\text{L}\cdot\text{kg}^{-1}$	0.39 ± 0.22	0.12 ± 0.04	0.148 ± 0.013	1.4 ± 0.6
F %	-	-	-	70 ± 3

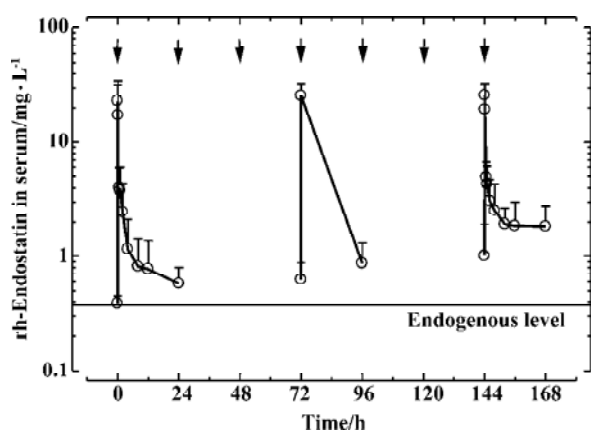


Figure 2. Concentration-time data of rh-endostatin after consecutive iv injection (q.d.×7) at a dose of 1.5 mg/kg in Rhesus monkeys. The endogenous endostatin levels set at 375±175 µg/L.

was 2.3±0.6 ($P=0.012$), indicating that the rh-endostatin had a tendency to accumulate in the body after successive iv administrations over 7 d with an interval of 24 h between doses.

Discussion

Folkman *et al* firstly described the angiogenesis inhibitor endostatin in January 1997^[2]. Although the molecular mechanisms behind the inhibition of angiogenesis have not yet been elucidated, endostatin seemed to inhibit endothelial cell migration *in vitro* and appeared to be highly effective in murine *in vivo* studies^[4]. Endostatin thought to be an ideal anticancer weapon, was quickly pushed into clinical trials^[5-8]. Thus far, however, recombinant endostatin prepared from *Escherichia coli* is insoluble after purification and therefore inappropriate for clinical settings. A soluble form of endostatin is available from a yeast system that has a relatively low yield and high cost, which has made it difficult to produce endostatin in quantities sufficient for extensive clinical evaluation^[9].

Researches attempted to solve the problems mentioned above, including using the His-tag technique^[10-12]. The rh-endostatin used in this study was His-tagged, containing a six-histidine residue sequence and an affinity tag at its N-terminal, so it could be effectively extracted by affinity chromatography based on immobilized metal ions such as Ni²⁺. In this capture step, we were able to purify 100% of the recombinant protein with more than 99% purity just as reported previously^[13]. The *in vivo* antiangiogenic and anticancer activities of this His-tag recombinant endostatin are as potent as those of the previously reported soluble form. Further, His-tag endostatin is also more convenient for analysis. In

our study, we utilized a rapid and sensitive approach, Matrix-Assisted Laser Desorption Ionization-Time of Flight-Mass Spectrometry (MALDI-TOF-MS), to analyze the proteins, which provided highly accurate the molecular weight information about the protein (data not shown).

There was a relatively high basic endostatin level in monkeys (375±175 µg/L), and it was consistent with endostatin being an endogenous protein. Moreover, to obtain an effect on tumors, the dose of endostatin in preclinical experiments and the recommended dose used in clinical trials are much higher than the dose required of most other therapeutic proteins, which reached the level of mg/kg (or mg/m²). Our PK studies showed that, although statistically not significant, λ_3 of the high dose group was obviously prolonged, suggesting that an exorbitant dose might affect the clearance of endostatin in the body. The results of the multiple dose study for 7 d continuously at the dose level of 1.5 mg/kg also suggested that rh-endostatin tended to accumulate in the body with a statistically significant accumulation factor of 2.3. Some clinical trials have shown that endostatin may cause side effects in the cardiovascular system, especially a dose-dependent toxicity in the heart. So it is best to avoid over accumulation of endostatin in the body by regulating the intervals between each administration. Furthermore, the PK study shows that differences between individuals are great, so individual monitoring during the course of treatment is also recommended in clinical trials.

Acknowledgment

The study was financially supported partly by MedGen Ltd (Yantai City, Shandong Province, China).

References

- 1 Sasaki T, Fukai N, Mann K, Gohring W, Olsen BR, Timpl R. Structure, function and tissue forms of the C-terminal globular domain of collagen XVIII containing the angiogenesis inhibitor endostatin. *EMBO J* 1998; 17: 4249-56.
- 2 O'Reilly MS, Boehm T, Shing Y, Fukai N, Vasios G, Lane WS, *et al*. Endostatin: an endogenous inhibitor of angiogenesis and tumor growth. *Cell* 1997; 88: 277-85.
- 3 Boehm T, Folkman J, Browder T, O'Reilly MS. Antiangiogenic therapy of experimental cancer does not induce acquired drug resistance. *Nature* 1997; 390: 404-7.
- 4 Zatterstrom UK, Felbor U, Fukai N, Olsen BR. Collagen XVIII/endostatin structure and functional role in angiogenesis. *Cell Struct Funct* 2000; 25: 97-101.
- 5 Iughetti P, Suzuki O, Godoi PH, Alves VA, Sertie AL, Zorick T, *et al*. A polymorphism in endostatin, an angiogenesis inhibitor, predisposes for the development of prostatic adenocarcinoma. *Cancer Res* 2001; 61: 7375-8.
- 6 Maio M, Altomonte M, Calabro L, Fonsatti E. Bioimmunotherapeutic

- targets on angiogenetic blood vessels in solid malignancies. *Front Biosci* 2001; 6: D776–84.
- 7 Ryan CJ, Wilding G. Angiogenesis inhibitors. New agents in cancer therapy. *Drugs Aging* 2000; 17: 249–55.
 - 8 Mundhenke C, Thomas JP, Wilding G, Lee FT, Kelze F, Chappell R, *et al*. Tissue examination to monitor antiangiogenic therapy: a phase I clinical trial with endostatin. *Clin Cancer Res* 2001; 7: 3366–74.
 - 9 Huang X, Wong MK, Zhao Q, Zhu Z, Wang KZ, Huang N, *et al*. Soluble recombinant endostatin purified from *Escherichia coli*: antiangiogenic activity and antitumor effect. *Cancer Res* 2001; 61: 478–81.
 - 10 Hengen P. Purification of His-Tag fusion proteins from *Escherichia coli*. *Trends Biochem Sci* 1995; 20: 285–6.
 - 11 Tallet B, Astier-Gin T, Castroviejo M, Santarelli X. One-step chromatographic purification procedure of a His-tag recombinant carboxyl half part of the HTLV-I surface envelope glycoprotein overexpressed in *Escherichia coli* as a secreted form. *J Chromatogr B Biomed Sci Appl* 2001; 753: 17–22.
 - 12 Gibert S, Bakalara N, Santarelli X. Three-step chromatographic purification procedure for the production of a his-tag recombinant kinesin overexpressed in *E. coli*. *J Chromatogr B Biomed Sci Appl* 2000; 737: 143–50.
 - 13 Huang YW, Lu ML, Qi H, Lin SX. Membrane-bound human 3beta-hydroxysteroid dehydrogenase: overexpression with His-tag using a baculovirus system and single-step purification. *Protein Expr Purif* 2000; 18: 169–74.
-

The 69th Annual Scientific Meeting of the Japanese Circulation Society

2005 Mar 19–21 Pacifico Yokohama, Japan

Please contact

Tetsu YAMAGUCHI, Chairman, MD, PhD,

Toranomon Hospital

2-2-2 Toranomon, Minato-ku, Tokyo 105-8470, Japan

Phn 81-3-3505-5872 Fax 81-3-3505-5873

E-Mail jcs69@convention.co.jp

URL <http://www2.convention.co.jp/jcs69/>